

ORIGINAL ARTICLE

Computer-aided design, synthesis, and characterization of molecular hybrids of dihydropyrazoles, aminopyrimidines, and thiazolidin-4-ones as potential inhibitors of the penicillin-binding protein 3 (PBP-3) of *Escherichia coli*

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Abstract

A computer-assisted approach was used to model and study privileged heterocyclic scaffolds containing dihydropyrazole, pyrimidin-2-amine, and thiazolidin-4-one moieties (hybrid pharmacophores) to obtain novel and promising antimicrobial prototype molecules. Main bioavailability descriptors were determined in order to assess the drug-likeness of the designed compounds and to pre-filter eleven compounds exhibiting the best profiles, thus passing to molecular docking study against a key penicillin-binding protein type-3 from enterotoxygenic *E. coli*. Seven structures were chosen by their energies of affinity and docking interactions with key residues in the active site of the receptor. Seven compounds with the highest docking scores belonging to the series of chalcones, dihydropyrazoles, aminopyrimidines, and thiazolidin-4-ones were prepared via condensation or cyclocondensation reactions. The structural elucidation of the final products was carried out by infrared spectra analysis and NMR experiments. Such molecular hybrids considered as potential hits in the search for new antibacterial compounds will be tested *in vitro* in further studies.

Keywords: PBP-3 inhibitors; hybrid pharmacophores; *in silico* screening; dihydropyrazoles; pyrimidin-2-amines; thiazolidin-4-ones.

1. Introduction

The inappropriate and indiscriminate application of antibiotics to treat dangerous and wellspread microbial infections has led to the evolution of multi-resistant and highly contagious bacterial strains responsible for high mortalities in immunocompromised patients and considerable morbidity in the community. Such bacteria include enterotoxigenic *Escherichia coli* (ETEC) and other multiresistant strains [1, 2]. ETEC is among the leading reported bacterial causes of diarrhea-related morbidity and mortality (around 157 000 deaths each year), mostly in children within developing countries, and it is also considered the bacterial strain responsible for traveler's diarrhea. Furthermore, the real burden of ETEC on public health is rather undervalued, because of the limited amount and availability of licensed vaccines for ETEC [3, 4, 5, 6].

This panorama has encouraged researchers to get involved in the discovery of new wide-spectrum antibacterial agents with low toxicity and high selectivity. Furthermore, their endeavors also strive for straightforward, resourceful, and cost-effective development protocols or synthetic pathways leading to health solutions, new knowledge, and innovative pharmacophores [7]. An



additional advantage of such efforts has been the boost of antibacterial drug-discovery research and industry, widely observed in the growing number of publications addressing the mentioned issues (selectivity and multi-drug resistance) and new antibiotics available today [7, 8, 9].

Antibacterial drug development is a lengthy and demanding process, whereby product safety and efficacy ought to be ascertained though clinical trials and therapeutics. Thus, it is paramount to make the whole process more rational by means of modern tools such as the SAR approaches (structure-activity relationships) and virtual screening [10, 11, 12].

Penicillin Binding Proteins (PBPs) synthesize and remodel peptidoglycan, the building block of bacterial cell walls giving the cell its shape and stiffness. PBPs are found in all bacteria and represent important targets for antimicrobial drug design. Along with PBPs, -lactamases are antimicrobial targets. These are of interest because of their high homology among different bacterial strains [13, 14].

PBP-3 (4BJP) of enterotixigenic *E. coli*, also known as FtsI, is one of the crucial enzymes involved in the final stages of peptidoglycan synthesis. PBP-3 is a homodimer made up of 588 amino acids in length, and it plays an essential role in cell division by catalyzing the peptides of cross-bridged peptidoglycan chains. Taking this into account, inhibition of PBP-3 will result in bacterial filamentation and death, thus becoming a particularly convenient target [15, 16]. In addition to the biological relevance of the ETEC's PBP-3 enzyme as a molecular target in antibiotic design, it is important to highlight that structure-based alignment studies of the PBP-3 transpeptidase domain exhibit really low structural divergence with respect to the same PBP-3 domain in the strains of *Acitenobacter baumanii*, *Pseudomonas aeruginosa*, and *Neisseria gonorrhoeae*, which could lead to the discovery of a broad spectrum antibiotic with potential activity against these additional and highly pathogenic resistant strains [17].

Compounds containing the thiazolidin-4-one core exhibit varied bioactivities as anticonvulsives [18], antileukemics [19], antibacterials [20, 21], and antitumorals [22], among others. The introduction of moieties with known biological activity, such as pyrimidine and pyrazole rings, is a good strategy in drug design, since those cores occupy an important place in heterocyclic chemistry as well as in medicinal chemistry, because of the wide range of chemical [23, 24], industrial and pharmacological properties they possess [25, 26, 27, 28]. The combination of such privileged heterocyclic scaffolds would circumvent the β -lactamase-based resistance mechanism, due to the absence of these four-membered rings in our molecular prototypes, which point to a competitive inhibition mechanism rather than a potential suicidal inhibition (general mechanism of action of β -lactam antibiotics) [14].

From this standpoint, and trying to make a contribution to this problem, we proposed the use of the widely studied and biologically promising *chalcones* as building blocks and the diversity oriented synthesis approach in the preparation of heterocyclic privileged scaffolds [29], molecular modification or hybrid pharmacophore [30], and bioisosterism [31], aiming at overcoming the multi-resistance mechanisms of bacteria and improving the bioavailability and bioactivity profiles of our potential antimicrobial drugs.

We undertook a computer-aided drug discovery approach (including ADMET predictions and virtual screening) with the goal of designing dihydropyrazole, pyrimidin-2-amine, and thiazolidin-4-one molecular hybrids as potential antimicrobial agents (via PBP-3 receptor inhibition) via synthetic transformation of chalcone precursors. With this strategy we aimed at improving our discovery stage in terms of bioactivity and atomic economy (lowering costs and waste generation).

2. Materials and Methods

2.1. Molecule modelling

Molecular modeling investigations were carried out using Dell XPS-8700 CPU running Intel[®] CoreTM i7-4790 Octacore Processor, 12 GB RAM, 2 TB hard disk and NVIDIA GeForce GTX 750Ti 2 GB graphics card. An *in silico* study was designed in order to screen the molecules exhibiting the best bioavailability, toxicity, and interaction with *E. coli* PBP-3 protein as molecular target. The most promising molecules were then selected to the synthesis phase. This study consisted of two stages as follows:

STAGE 1. ADMET properties determination

Initially, all molecules were analyzed using the Medchem DesignerTM software version 3.0.030, Copyright[©] 2011-2014, Simulations Plus, Inc. MedChem Designer uses innovative molecule drawing features to provide free, fast, and accurate Absorption, Distribution, Metabolism, Excretion and Toxicity (ADMET) predictions, based on the ADMET PredictorTM database and Lipinski's Rule of five. When using this software, the following properties are obtained: $M \log P$, $S + \log P$, and $S + \log D$. These constitute descriptors of the partition coefficient, namely the rule of five as the number of Lipinski's Rules broken by the molecules. The RuleOf5_Code employed entails the codes for rules being broken, M.W. as the molecular weight of the molecules, M_NO as the number of hydrogen bond aceptors, T_PSA as the polar surface area, and HBDH as the number of hydrogen bond donors.

The molecules breaking none, or at most one, of Lipinski's Rules were then analyzed with the online server admetSAR, an ADMET structure-activity relationship server that predicts ADMET properties of drug candidates and environmental chemicals [32]. The admetSAR tool can predict the classification of the molecules according to their ADMET features, giving a probability for each classification.

From the information obtained employing admetSAR for the molecules that had passed bioavailability filter, the AMES toxicity classification and the Rat Acute Toxicity regression results were further used to choose the molecules for the molecular docking stage.

STAGE 2. Molecular docking

Molecular docking was carried out following three steps:

Step 1. Structure optimization. The molecules were optimized using the software HyperChemTM 8.0.10 for Windows[©] 1995-2011, Hypercube Inc. The molecular mechanics protocol consisted of an MM+ force field, optimizing until obtaining an RMS gradient equal to or below 0.01 kcal mol⁻¹, using a Polak-Riviere algorithm of conjugated gradients. The idea with this process was to obtain the lowest energy conformations of each compound and have them ready for the docking process.

Step 2. Protein preparation. Before beginning the docking process, the molecular target was prepared. The PBP-3 structure was downloaded from Protein Data Bank (PDB) under the code 4bjp [15], and prepared employing PyMOL Molecular Graphics System version $2.3.2_0^{\circ}$ 2003-2019, Schrödinger LCC and Autodock Tools version 4.2.6 Copyright [©] 1999-2014, as follows: Removal of all bound water molecules, ligands and co-factors (preprocess) from the protein; assignment of bond orders; addition of hydrogens (polar only); optimization of hydrogen bonds; and correction of charges.

Step 3. Molecular docking. The molecules that passed through phases 1 and 2, along with *Ampicillin* (our reference or blank) were docked against one monomer of PBP-3 receptor of *E. coli.* For this step, the program Autodock Tools was used (version 4.2.6 Copyright[©] 1999-2014, Molecular Graphics Laboratory – The Scripps Research Institute). The docking protocol was validated before conducting the docking studies by reproducing the natural bound ligand conformation of 3-cyclohexyl-1-propylsulfonic acid (CXS) in the crystal structure of PBP-3 (PDB ID: 4bjp). The co-crystal (CXS) was removed of the catalytic pocket and re-docked, while the best three docking poses were considered in order to validate the results by calculating root-mean-square deviation of atomic positions – RMSD of such configurations in comparison to the co-crystallized ligand (RMSD in the range 1.15 Å to 1.82 Å). The top 10 conformations for each molecule were analyzed using PyMOL and Discovery Studio Visualizer[©] (version 16.1.0.15350 2015, Dassault Systèmes Biovia Corp). The molecules that could interact with one or more amino acids of each domain, being at least one of those crucial for the interaction of the domains and ligand recognition, were selected for the synthesis stage.

2.2. Molecule synthesis

The material used for molecule synthesis was dried in an oven at 180 °C for 24 hours before use. The raw materials, products and reagents were weighed in the Wisd WBA-220 analytical balance. The reactions were conducted in a heating and stirring plate Wisd dh DAITHAN Scientific Maxtir TM 500. The concentration of the reaction products was performed in a rotary evaporator IKA RV10 equipped with a vacuum pump MPC105T IKA version.

Purification of the synthesized compounds was carried out by column chromatography using 230-240 mesh Macherey-Nagel silica gel (0.040 mm to 0.063 mm), fractional distillation, and fractional recrystallization as required. The melting points of the raw materials and products were determined on a Stuart SMP10 Facsia 120v melting point apparatus (Cole-Parmer Ltd, United Kingdom). Chromatographic control of all reactions raw materials and the products was performed by Thin-layer chromatography (TLC), using 20 cm \times 20 cm aluminum chromatoplates coated with 60 F₂₅₄ silica gel (Merck, Germany). A UVP Spectroline Longlife TM Filter (SW 254 nm and LW 365 nm) UV lamp was used for the development of the TLC plates.

Infrared (IR) spectra were determined on an IR-Nicolet iS10 spectrometer (Thermo Fisher Scientific, USA) with KBr pellet in the range 4000 cm^{-1} to 500 cm^{-1} . Nuclear magnetic resonance spectra of all the obtained compounds were determined on a Bruker Advance 400 spectrometer (Bruker, USA). For ¹H at 270 MHz and for ¹³C to 67.89 MHz spectra; and in a Varian model VNMRS-500 for ¹H at 500 MHz and for ¹³C at 125.787 MHz. This determination was carried out using deuterated chloroform (CDCl₃) and deuterated dimethylsulfoxide (DMSO-d₆) and tetramethylsilane (TMS) as internal reference. The elemental analyses were carried out on a LECO CHNS-900 elemental analyzer (LECO corporation, USA).

2.3. Synthesis of 3-(4-methoxyphenyl)-1-(4-methylphenyl)prop-2-en-1-one (compound 1h)

The synthesis of this chalcone was carried out mixing 4-methoxybenzaldehyde (1 eq) and 4-methylacetophenone (1.1 eq) in the presence of industrial ethanol (20 mL) in a balloon. The mix was stirred at a temperature of 60 °C to improve the solubility of the reagents. After the formation of a beige precipitate, the balloon was placed in an ice bath for 15 min until abundant solid was observed, the solid was then vacuum-filtered and washed with cold ethanol to yield the corresponding chalcone (82 % yield).

Synthesis of 3-(4-methoxyphenyl)-1-(4-methylphenyl)prop-2-en-1-one (1 h). [33] Beige solid, 82 % yield, mp: 63 °C to 65 °C. ¹H NMR (400 MHz, CDCl₃), $\delta_{\rm H}$: 7.93 (d, J = 8.20 Hz, 2H), 7.78 (d, J = 15.63 Hz, 1H), 7.60 (d, J = 8.78 Hz,2H), 7.42 (d, J = 15.61 Hz, 1H), 7.30 (d, J = 8.06 Hz, 2H), 6.94 (d, J = 8.78 Hz, 2H), 3.85 (s, 3H), 2.43 (s, 3H). ¹³C NMR (100 MHz, CDCl₃), $\delta_{\rm C}$: 190.2, 161.7, 144.4, 143.5, 136.1, 130.3, 129.4, 128.7, 127.9, 119.9, 114.5, 55.6, 21.8.

2.4. Synthesis of 1,3,5-triaryl-4,5-dihydro-1*H*-pyrazolines (compounds 2b and 2c)

The precursor of compound 2b: 3-(4-ethoxyphenyl)-1-(4-methylphenyl)prop-2-en-1-one and the precursor of compound 2c: <math>3-(4-ethoxyphenyl)-1-phenylprop-2-en-1-one (1eq) were placed each in a round-bottomed flask and suspended in ethanol (10mL), with an excess of phenylhydrazine (2eq). The mixtures were allowed to react under reflux for four hours until the formation of a light-yellow solution, which was then evaporated under reduced pressure. The resulting reaction crudes were dissolved in CHCl₂ (4 mL), extracted with a saturated solution of sodium bicarbonate (3 portions, 5 mL each one), and dried with anhydrous NaSO₄. Each extract was evaporated under reduced pressure, and the residues were recrystallized from methanol yielding the corresponding dihydropyrazole derivative (87 % and 85 percent yield, respectively).

5-(4-ethoxyphenyl)-3-(4-methylphenyl)-1-phenyl-4,5-dihydro-1*H***-pyrazole (compound 2b).** Light yellow solid, 87 % yield, mp: 153 °C to 154 °C. ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 7.63 (d, J = 8.14 Hz, 2H), 7.26-7.17 (m, 6H), 7.09 (d, J = 8 Hz, 2H), 6.86 (d, J = 8.67 Hz, 2H), 6.78 (t, J = 8 Hz, 1H), 5.20 (dd, $J_{\rm cis} = 7.25$ Hz, $J_{\rm trans} = 12.19$ *Hz*, 1H), 4.00 (q, J = 6.99*Hz*, 2H), 3.79 (dd, $J_{\rm trans} = 12.25$ *Hz*, $J_{\rm gem} = 17.05$ Hz, 1H), 3.10 (dd, $J_{\rm cis} = 7.27$ Hz, $J_{\rm gem} = 17.05$ Hz, 1H), 2.39 (s, 3H), 1.40 (t, J = 6.98 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) $\delta_{\rm C}$: 158.4, 147.1, 145.2, 138.7, 134.7, 130.2, 129.4, 128.9, 127.2, 125.8, 118.9, 115.1, 113.5, 64.1, 63.5, 43.9, 21.5, 14.9. Anal. Calcd for C₂₄H₂₄N₂O: C, 80.87; H, 6.79; N, 7.86. Found: C, 80.79; H, 6.71; N, 7.78.

5-(4-ethoxyphenyl)-1,3-diphenyl-4,5-dihydro-1*H***-pyrazole (2c).** Light yellow solid, 85 % yield, mp: 93 °C to 95 °C. ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 7.74 (d, 2H), 7.40 (t, 2H), 7.36 - 7.33 (m, 1H), 7.26 - 7.18 (m, 4H), 7.10 (d, 2H), 6.86 (d, 2H), 6.80 (t, 1H), 5.23 (dd, $J_{\rm cis} = 7.22$ Hz, $J_{\rm trans} = 12.27$ Hz, 1H), 4.00 (q, J = 7.01 Hz, 2H), 3.81 (dd, $J_{\rm trans} = 12.30$ Hz, $J_{\rm gem} = 17.06$ Hz, 1H), 3.12 (dd, $J_{\rm cis} = 7.23$ Hz, $J_{\rm gem} = 17.06$ Hz, 1H), 1.40 (t, J = 6.98 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) $\delta_{\rm C}$: 158.5, 146.8, 145.0, 134.6, 132.9, 128.9, 128.7, 127.2, 125.8, 119.1, 115.1, 113.5, 64.2, 63.5, 43.7, 14.9. Anal. Calcd for C₂₃H₂₂N₂O: C, 80.67; H, 6.48; N, 8.18. Found: C, 80.73; H, 6.41; N, 8.22.

2.5. Synthesis of 4,6-diaryl-2-aminopyrimidines (compounds 3a and 3h)

The synthesis of compounds 3a and 3h involved: 10 mmol of the proper aromatic aldehyde, 10 mmol of the corresponding substituted acetophenone, 16 mmol of guanidine hydrochloride, and 20 mmol of NaOH. The reactants were mixed in 20 mL of EtOH and refluxed for 12 hours under nitrogen atmosphere. After completion of the reaction, 15 mL of diluted hydrochloric acid was added to the crude, giving rise to an emulsion that was then destroyed by extraction with DCM (3 portions, 15 mL each one). The resulting organic layer was dried with anhydrous MgSO₄ and evaporated under reduced pressure, yielding the final products 3a and 3h (orange solid) which were left to dry in oven at 70 °C for five hours for a final yield of 85 % and 69 %, respectively.

4-(2-chlorophenyl)-6-(4-methylphenyl)pyrimidin-2-amine (3a). Orange solid, 85 % yield, mp: 62 °C to 63 °C. ¹H NMR (400 MHz, CDCl₃) δ_{H} : 7.93 (d, J = 8.22 Hz, 2H), 7.61 - 7.59 (m, 1H), 7.50 - 7.48 (m, 1H), 7.38 - 7.35 (m, 2H), 7.34 (s, 1H), 7.27 (d, J = 7.95 Hz, 2H), 5.47 (s, 2H), 2.41 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ_{C} : 165.9, 165.6, 163.6, 141.0, 138.0, 134.8, 132.3, 130.9, 130.4, 130.3, 129.6, 127.3, 127.2, 108.4, 21.5. Anal. Calcd for C₁₇H₁₄ClN₃: C, 69.03; H, 4.77; N, 14.21. Found: C, 68.97; H, 4.81; N, 14.27.

4-(3-methoxyphenyl)-6-(4-methylphenyl)pyrimidin-2-amine (3h). Orange solid, 69 % yield, mp: 119 °C to 121 °C. ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$:8.02 (d, 2H), 7.98 (d, 2H), 7.42 (s, 1H), 7.32 (d, 2H), 7.01 (d, 2H), 5.25 (s, 2H), 3.94 (s, 3H), 2.44 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) $\delta_{\rm C}$: 165.9, 165.6, 163.6, 161.0, 140.4, 137.6, 130.1, 129.5, 128.6, 126.6, 114.6, 103.2, 51.4, 21.9. Anal. Calcd for C₁₈H₁₇N₃O: C, 74.20; H, 5.88; N, 14.42. Found: C, 74.17; H, 5.81; N, 14.37.

2.6. Synthesis of 3-[4,6-(diaryl)pyrimidin-2-yl]- 2-phenyl-1,3-thiazolidin-4-one hybrids (compounds 4g and 4h).

In a round-bottomed flask, 1.0 mmol of the corresponding aminopyrimidine were dissolved in 20 mL of dry benzene, with 3.0 mmol of 2-mercaptoacetic acid and 1.5 mmol of the benzaldehyde with stirring. The mixture was refluxed for 8 hours with a Dean-Stark trap and the reaction was monitored by TLC. After completion, the resulting crude was cooled to room temperature and neutralized with a 10 % sodium bicarbonate solution (4 portions of 10 mL). The organic phase was dried with anhydrous sodium sulphate, evaporated under reduced pressure, and the resulting crude was dissolved by heating in absolute ethanol. After cooling to room temperature, the final product obtained as precipitate was allowed to dry for 24 hours to yield two pyrimidinylthiazolidin-4-one hybrids (43 % to 48 % yield).

3-[4-(3-ethoxyphenyl)-6-(4-methylphenyl)pyrimidin-2-yl]-2-phenyl-1,3-thiazolidin-4-one (4g). Light yellow solid, 43 % yield, mp: 102 °C to 104 °C. ¹H NMR (400 Hz, DMSO-d₆) $\delta_{\rm H}$: 8.15 (d, J = 7.75 Hz, 3H), 7.76 (d, 1H), 7.70 (s, 1H), 7.46 (d, J = 7.49 Hz, 2H), 7.34 - 7.27 (m, 4H), 7.20 (t, 1H), 6.90 - 6.85 (m, 2H), 4.11 - 4.05 (m, 3H), 3.92 (d, J = 16.09 Hz, 1H), 2.38 (s, 3H), 1.39 (t, 3H). ¹³C NMR (100 MHz, DMSO-d6) $\delta_{\rm C}$: 170.6, 165.1, 164.0, 162.3, 159.6, 141.5, 139.1, 138.0, 136.4, 132.6, 129.1, 128.6, 127.8, 127.6, 126.0, 121.8, 116.3, 112.7, 108.7, 63.6, 62.5, 34.5, 22.8, 14.8. Anal. Calcd for C₂₈H₂₅N₃O₂S: C, 71.92; H, 5.39; N, 8.99; S, 6.86. Found: C, 71.88; H, 5.33; N, 9.07; S, 6.79.

3-[4-(4-methoxyphenyl)-6-(4-methylphenyl)pyrimidin-2-yl]-2-phenyl-1,3-thiazolidin-4-one (4h). Light yellow solid, 48 % yield, mp: 80 °C to 82 °C. ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 8.00 (d, J = 6.85 Hz, 2H), 7.90 (d, J = 6.50 Hz, 2H), 7.70 (s, 1H), 7.45 (d, J = 7.36 Hz, 2H), 7.30 - 7.21 (m, 5H), 6.81 (s, 1H), 6.97 (d, J = 6.52 Hz, 2H), 4.07 (d, 1H; J = 16.11 Hz), 3.89 - 3.86 (m, 4H), 2.41 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) $\delta_{\rm C}$: 170.9, 165.6, 165.4, 162.3, 157.6, 141.5, 141.5, 134.1, 129.7, 129.3, 129.1, 128.9, 128.3, 127.4, 126.1, 114.4, 107.8, 63.8, 55.6, 34.3, 21.6. Anal. Calcd for C₂₇H₂₃N₃O₂S: C, 71.50; H, 5.11; N, 9.26; S, 7.07. Found: C, 71.47; H, 5.20; N, 9.19; S, 7.11.

3. Results and Discussion

3.1. Virtual Screening

We identified key biologically promising units and modelled their structures following a hybrid pharmacophore strategy, involving the combination of privileged scaffolds with renowned antibacterial activity (chalcones, β -lactam rings, dihydropyrazoles, pyrimidin-2-amines, and thiazolidin-4-ones). In order to determine Lipinski-based bioavailability and drug-likeness properties [34], 72 new molecular prototypes were proposed and screened *in silico* by their ADMET profiles, introducing structural diversity through their substituents (**Table 1**).

The computational prediction of ADMET properties for such 72 proposed compounds, was followed by a complete study of their drug-like behavior, allowing us to choose 11 prototypes to move forward. As observed in **Table 2**, five of the best 11 compounds violated only the lipophilicity parameter, whereas compounds 1h, 3a, 3h, 4g, 4h, and 5l exhibited no violations of Lipinski's rule of five. The 11 compounds selected presented an ADMET profile and drug-likeness comparable to that of the reference drug Ampicillin, making them interesting in terms of bioavailability.

Having studied the bioavailability of the 11 selected prototypes, we proceeded with an *in silico* toxicity analysis of such compounds. We applied a structure-activity relationship tool empowered by a complete database of drugs and different chemicals [32], known as admetSAR software. The toxicity predictions revealed that derivatives with $-NO_2$ substituents were potential carcinogens and likely threats to the environment; whereas most of compounds with a -OH functional group in their aromatic rings were discarded for their potential generation of toxic metabolites (as

R I	Ph R' R R' R' 2	R ^{NH2} R ^N R ^I 3	$ \begin{array}{c} $	$rac{Ph}{R}$	CI R 6
Entry	-R	-R'	Entry	-R	-R'
a	Me	CI	g	Me	€ €
b	<u>Қ</u> Ме	ξ− √ −OEt	h	<u></u> Ме	§ОМе
с		€OEt	i		<u></u> Ме
d		ξ-NO ₂	j	Me	δH −OEt
e	OH	NO ₂	k	OH	ξ− ∕ −OMe
f	OH	€ OH OEt	1	ễ− Me	

Table 1: Structures of the proposed molecular hybrids and their substitution pattern 1-6(a-l).

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Compound	MlogP	S+logP	RuleOf5	RuleOf5_Code	M.W.	M_NO	T_PSA	HBDH
1h	3.641	4.259	0	<none></none>	252.32	2	26.3	0
2b	4.909	5.732	1	LP*	356.47	3	24.83	0
2c	4.698	5.233	1	LP*	342.43	3	24.83	0
3 a	4.329	4.452	0	<none></none>	295.77	3	51.8	2
3h	3.521	4.105	0	<none></none>	291.36	4	61.03	2
4 g	3.773	4.387	0	<none></none>	467.59	5	55.32	0
4h	4.093	4.343	0	<none></none>	453.57	5	55.32	0
5j	4.642	4.754	1	LP*	446.57	5	53.01	1
51	3.678	2.831	0	<none></none>	310.42	3	23.55	0
6a	6.027	4.805	1	LP*	409.32	3	23.55	0
6c	5.424	5.033	1	LP*	418.93	4	32.78	0
Ampicillin	0.656	-1.302	0	<none></none>	349.41	7	112.73	4

Table 2: Predicted bioavailability profiles of the group of 11 molecular hybrids and *Ampicillin*. *LP = Lipophilicity.

suggested by admetSAR's CYP metabolite analysis). In comparison to *Ampicillin*, the LD_{50} values observed for the majority of the molecules turned out similar, but close to the edge of the Hodge toxicity scale [34]. Compound 5j exhibited a lower toxicity score in comparison to the remaining prospects (**Table 3**), but, then again due to the presence of hydroxylated aromatic ring as substituent, the possibility of production of toxic metabolites could not be ruled out.

The eleven structures exhibiting the best ADMET scores were energy-minimized with the HyperChem v.8.0.10 software, yielding the most stable molecular conformation for each prototype and the proper files of the ligands for the docking protocols (**Table 4**). Such optimization allowed us to observe that nine of the structures presented lower conformational energies than that of Ampicillin. Compounds 6a and 6c had energies similar to the reference drug, and compounds 5j and 5l exhibited higher energy values than Ampicillin. The latter two structures might be conformationally unstable or more reactive.

We studied the viability of the interaction of the selected molecules with the catalytic domain of the enzyme transpeptidase PBP-3 (PDB ID: 4bjp) of enterotoxigenic *E. coli* and identified the most likely ligand-receptor binding conformation via computer molecular coupling analysis. The study of molecular docking was conducted by means of a rigid-body approach and the study was

Compound	Rat Acute Toxicity LD50, $mg kg^{-1}$	Fish Toxicity $pLC50$, mg L ⁻¹	Tetrahymena Pyriformis Toxicity pIGC50, $\mu g L^{-1}$	AMES Toxicity	Carcinogens
1h	457.2	0.2367	1.9444	Non AMES toxic	Non-carcinogen
2b	777.6	0.6985	0.9715	Non AMES toxic	Non-carcinogens
2c	741.12	0.7363	0.9568	Non AMES toxic	Non-carcinogens
3a	611.98	0.9897	1.0663	Non AMES toxic	Non-carcinogens
3h	555.09	2.1895	0.4607	Non AMES toxic	Non-carcinogens
4g	1079.17	1.4308	0.5852	Non AMES toxic	Non-carcinogens
4h	975.35	1.366	0.5324	Non AMES toxic	Non-carcinogens
5j	1107.54	1.149	0.591	Non AMES toxic	Non-carcinogens
51	760.65	1.7126	0.2589	Non AMES toxic	Non-carcinogens
6a	977.08	0.9791	0.9481	Non AMES toxic	Non-carcinogens
6c	1000.56	0.8618	0.8056	Non AMES toxic	Non-carcinogens
Ampicillin	545.78	1.7369	0.2178	Non AMES toxic	Non-carcinogens

Table 3: Toxicity profiles of the selected eleven molecular hybrids and Ampicillin.

Compound	Energy $(kJ mol^{-1})$	Energy Gradient (kJ mol ⁻¹)
1h	67.61	0.03724
2b	37.36	0.03891
2c	38.62	0.03766
3 a	48.58	0.04142
3h	41.38	0.04017
4 g	38.99	0.03975
4h	34.43	0.04058
5ј	119.29	0.03682
51	106.82	0.041
6a	370.12	0.03891
6c	375.81	0.04017
Ampicillin	369.45	0.03891

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Table 4: Molecular c	optimization energies an	d gradients of the selected	d eleven molecular hybrids.
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performed at the active site identified for the enzyme PBP-3, taking into consideration eight key amino acid residues involved in ligand recognition: Ser307, Lys310, Ser359, Asn361, Lys494, Thr495, Gly496, and Thr497, responsible for the binding of β -lactams to the active site of PBP's.

The different docking results (**Table 5**) revealed which of the selected molecules display binding energy values similar to that of the reference drug *Ampicillin* ($-6.16 \text{ kJ mol}^{-1}$), and which of them have a better interaction with the amino acids of the catalytic site of the enzyme. We found that prototypes 4g, 4h, 2b, and 2c presented the best affinities in terms of binding energies and interactions (hydrogen bonds mainly) with several amino acids (most of them crucial) at the catalytic pocket, taking into consideration which of their atoms participate in those binding interactions (Table 5). Also, prototypes 5j and 5l had no apparent interaction with the enzyme's active site; whereas, pyrimidines 3a and 3h exhibited lower binding energies.

 Table 5: Binding energies and interactions of the eleven selected prototypes and Ampicillin with active site residues.

Prototype	Structures	Binding energy (kg mol ⁻¹)	Hydroge (residue-in	
1h	Me	-5.78	SER 307:OH THR 497:NH TYR 347:OH	1b:9:OMe 1b:18:O=C 1b:18:O=C
2b	Me	-6.26	SER 307:OH SER 307:OH	2b:1:NPh 2b:2:N=C

2c	NN OEt	-6.22	SER 307:OH 2c:1:N=C ALA 544:NH 2c:18:OEt SER 359:OH 2c:1:N=C
3a	CI N N Me	-5.43	PHE 417:O 3a:31:HN ASN 361:O 3a:32:HN TYR 419:OH 3a:2:N=C
3h	Me OMe	-5.56	ASN 361:O 3h:34:HN TYR 419:OH 3h:21:OMe PHE 417:O 3h:34:HN
	Me OEt	-6.79	ASN 361:NH 4g:1:S TYR 514:OH 4g:6:O=C SER 307:OH 4g:6:O=C THR 497:OH 4g:6:O=C
4h	Me OMe	-6.87	ASN 361:NH 4h:1:S SER 307:OH 4h:4:N-C TYR 511:OH 4h:12:N=C LYS 499:NH 4h:31:OMe ALA 544:NH 4h:31:OMe GLY 496:OH 4h:6:O=C THR 497:OH 4h:6:O=C
5j	S S O H O Et	-5.58	LYS 499:NH 5j:44:OH
51	o N.N.N.	-5.69	NO
6a		-4.52	SER 307:NH 6a:37:O=C SER 359:NH 6a:37:O=C
6c	CI EtO	-5.23	THR 497:OH 6c:19:OEt
Ampicillin	H ₂ N O O O O O O H	-6.16	ASN 361:0 AMP:42:OH SER 397:NH AMP:24:O=C ALA 544:NH AMP:11:O=C ALA 544:NH AMP:11:O=C SER 359:NH AMP:24:O=C SER 359:O AMP:43:NH



Figure 1: Binding conformation of molecular hybrid 4h with the active site of the PBP3 receptor of *E. coli*: a) Main interactions with crucial amino acid residues. b) Docking pose of compound 4h inside the catalytic pocket (Images generated with Pymol viewer).

After the enzyme-ligand molecular coupling study, each prototype was visualized and analyzed by means of the PyMOL computational tool (**Figure 1**), which determined that ten compounds exhibited interaction with at least one of the key amino acid residues present at the active site of *E. coli* PBP-3. Among the compounds that showed interaction with the catalytic site of the enzyme, one is a *chalcone* (1h), two are molecules with dihydropyrazole nuclei (2b, 2c), two aminopyrimidine derivatives (3a, 3h), two hybrids pyrimidinylthiazolidin-4-ones (4g, 4h), and two hybrid derivatives with fused pyrazole and -lactam nuclei (6a, 6c).

According to the binding energies and interactions exhibited by the mentioned molecular prototypes, hybrid compounds 5j (interaction with a secondary and less important residue) and 5l (no interaction with key residues of active site) were discarded due to their low profile of bioactivity *in silico*. Candidates 6a and 6c were also discarded because of their low binding energies, as well as poor polar interactions (two and one interactions respectively) with the aforementioned key amino acid residues.

From our complete docking analysis, molecular hybrids 4g and 4h could be considered as the most promising in terms of their affinity (binding energy and interactions with key amino acid residues) with the active site of PBP-3, thus becoming two potential hits in the search for a new inhibitor of the molecular target (Figure 1 and **Figure 2**).

Figure 1 and Figure 2 illustrate the docking results of the molecular hybrid 4h with the enzyme PBP-3, which exhibited a binding energy of $-6.87 \text{ kJ} \text{ mol}^{-1}$ and key interactions with the active site (Table 5), with five amino acids of the receptor (ASN 361, SER 307, LYS499, ALA544, GLY 496, and THR 497). Among these residues, ASN 361, SER 307, GLY 496, and THR 497 are crucial within the active site of the catalytic pocket. The main interactions observed occurred between the oxygen of the C=O group of the thiazolidin-4-one ring and the OH group of residues THR 497 at 1.9 Å and GLY 496 at 2.1 Å. Also, the –OMe substituent of ring B interacted with the NH group of ALA 544 at 2.2 Å or with residue LYS 499 at 2.3 Å. Other interactions were observed involving the heteroaromatic N-atom of the pyrimidine ring with the OH group of TYR 511 at 2.1 Å, and the N-atom of thiazolidin-4-one ring with the –OH group of SER 307 at 2.4 Å. A polar interaction (non-conventional hydrogen bond) was detected between the NH group of the residue ASN 361 and the sulfur atom of the thiazolidin-4-one ring at 2.2 Å. Additionally, other



Figure 2: Binding mode analysis of molecular hybrid 4h with the active site of PBP-3 of *E. coli* (drawing based on DS visualizer 2D diagram).

non-covalent interactions such as $\pi - \pi$ stacking interactions were observed, involving benzene rings A and B, as well as the pyrimidine ring with some aminoacidic residues containing aromatic rings (TRP 405, TYR 419 and TYR 511, PHE 415, and PHE 526).

3.2. Synthesis stage of the selected compounds

The synthesis of the selected compounds was carried out starting from an array of different precursors. The first precursor type consisted of , -unsaturated carbonyl compounds (chalcones, prototype 1). Such precursor was prepared via the Claisen-Schmidt type cyclocondensation reaction between aromatic aldehydes and the corresponding substituted acetophenones, according to the procedure reported in the literature [35]. On the other hand, dihydropyrazole derivatives (prototype 2) were obtained by cyclocondensation reactions from the reaction of the mentioned chalcones with one equivalent of arylhydrazines. Substituted aminopyrimidine series (prototype 3) were prepared allowing the reaction of chalcones with guanidine hydrochloride in basic medium, thereby obtaining the named derivatives (**Table 6**). Both, dihydropyrazoles and aminopyrimidines exhibited high fluorescence properties in solution, making them suitable for applications in molecular materials science.

Lastly, thiazolidine-4-ones (Prototype 4) were obtained from the aminopyrimidines described above, by means of a three-component cyclocondensation reaction with 2-mercaptoacetic acid and benzaldehyde in anhydrous benzene as solvent, using conditions of reflux and azeotropic removal of water with Dean- Stark trap (Table 6).



Table 6: General synthesis scheme and experimental data of the final eight selected compounds belonging to prototypes 1-4.

Most of the final products were synthesized in good yields and at reasonable reaction times by means of relatively simple experimental protocols. Prototypes 1-3 were synthesized and reported previously [36, 37], but compounds 4g and 4h, even though isolated in moderate yields, were prepared and reported for the first time in the present work. All of the obtained compounds were fully characterized and confirmed by IR spectroscopy, ¹H-NMR, ¹³C-NMR, and two-dimensional techniques.

All seven selected compounds were successfully synthesized and isolated as pure products, allowing a full characterization by detailed NMR and Elemental Analysis (EA) measurements (see Materials and Methods and **Suppl. 1**). An interesting feature of the ¹H NMR spectra of dihydropyrazoles 2b and 2c revealed that the two methylene H-4 protons and H-5 proton at the chiral center of the pyrazoline moiety form together an ABX spin system. Thus, the H_A-4 and H_B-4 appear each as a double-doublet at $\delta = 2.75$ ppm and 3.14 ppm, and $\delta = 3.11$ ppm and 3.38 ppm, respectively, with ²J_{AB} = 7.0 Hz to 13.6 Hz; whereas, H_X-5 appears as a double-doublet at $\delta = 5.25$ ppm to 5.15 ppm with ³J_{AX} = 7.0 Hz to 8.0 Hz and ³J_{BX} = 4.080Hx for both

compounds. The main signals in the ¹³C NMR spectra for compounds 2b and 2c correspond to C-4 at $\delta = 40.9$ ppm and 43.8 ppm, C-5 at $\delta = 59.4$ ppm for both compounds, and the quaternary C-3 at $\delta = 152.5$ ppm also for both derivatives.

As for the aminopyrimidines 3a and 3i, the protons of their NH_2 group display a characteristic broad singlet at 5.50 ppm and 5.21 ppm, respectively; whereas pyrimidine proton H-5 appears at 7.34 ppm for 3a and 7.41 ppm for 3i, differing slightly from data reported previously in the literature [35, 36]. The ¹³C NMR and DEPT-135 spectra of 3a and 3i exhibited the expected number and type of signals, allowing us to fully characterize those compounds.

Similar to the previously described dihydropyrazoles 2b and 2c, the molecular hybrids 4g and 4h presented diastereotopic protons at their methylene position H-5 ($\delta = 2.90$ ppm to 3.90 ppm), by long-range coupling via sulfur atoms with the H-2 proton ($\delta = 4.11$ ppm to 4.21 ppm) at the chiral carbon of the thiazolidin-4-one ring.

Finally, the spectroscopic data of compound 1h agreed with a previous report [33], and the purity of compounds 2-4 was confirmed by means of EA, where the corresponding elemental composition (CHNX) was measured and compared to the calculated value in all cases.

4. Conclusions

Following a methodology based on computer-aided drug design, it was possible to propose and study a series molecular prototypes conformed by privileged scaffolds such as chalcones, aminopyrimidines, thiazolidin-4-ones, pyrazoles, and β -lactams.

Seven of these molecules, exhibited interesting binding affinities in terms of energy and interactions with some of the key amino acids that make up the active site of PBP3 (4BJP) of enterotoxigenic *E. coli*, thus these are considered as potential hits in the search for new antibiotics.

The seven selected prototypes were synthesized and isolated in moderate to good yields and can be considered promising candidates for subsequent *in vitro* tests in further studies, aiming at endorsing their bioavailability profiles and possible antimicrobial activities, as a step beyond the computational tests performed in the present study.

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6. Conflict of interest

The authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest (such as honoraria; educational grants; participation in speaker membership, employment, consultancies, stock ownership, or other equity interest; and expert testimony or patent arrangements), or non (such as personal or professional relationships, affiliations, knowledge or beliefs) in the subject matter or materials discussed in this manuscript. We wish to confirm that there are no known potential conflicts of interest related to individual authors' commitments, or potential conflicts of interest related to commitments of editors, journal staff, or reviewers.

References

- Cuellar LE, Infecciones en huéspedes inmunocomprometidos. *Revista Medica Herediana*, 24: 156-161, 2013.
- [2] Bourgeois AL, Wierzba TF, Walker RI. (2016). Status of vaccine research and development for enterotoxigenic Escherichia coli. *Vaccine*. 34 (26): 2880–2886, 2016.

doi: 10.1016/j.vaccine.2016.02.076

[3] Gupta SK, Keck J, Ram PK, Crump JA, Miller MA, Mintz ED. Analysis of Data Gaps Pertaining to Enterotoxigenic Escherichia coli Infections in Low and Medium Human Development Index Countries, 1984-2005. *Epidemiology and Infection*. 136 (6): 721–738, 2008.

doi: 10.1017/S095026880700934X

[4] Kotloff KL, Nataro JP, Blackwelder WC, et al. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. *The Lancet.* 382 (9888): 209–222, 2013.

doi: 10.1016/s0140-6736(13)60844-2

[5] Ochoa TJ, Contreras CA. Enteropathogenic Escherichia coli infection in children. *Current Opinion in Infectious Diseases*. 24: 478–483, 2011.

doi: 10.1097/QCO.0b013e32834a8b8b

[6] Contreras CA, Ochoa TJ, Lacher DW, et al. Allelic variability of critical virulence genes (eae, bfpA and perA) in typical and atypical enteropathogenic Escherichia coli in Peruvian children. J. Med Microbiol. 59: 25–31, 2010.

doi: 10.1099/jmm.0.013706-0

- [7] World Health Organization. (2016). Antibiotic resistance. Fact sheet.
- [8] Vardanyan R, Hruby V. Synthesis of Best-Seller Drug. University of Arizona, Academic Press, Tucson, AZ, pp 847, 2016.
- [9] Reguero M, Barreto E, Jimenez F. Relacion estructura quimica actividad biológica. Una revisión retrospectiva. *Revista Colombiana de de Ciencias Quimico-farmaceuticas*, 17: 81-84, 1989.

doi: 10.15446/rcciquifa

- [10] Amábile-Cuevas C. In many ways, the fight against antibiotic resistance is already lost; preventing bacterial disease requires thoughtful new approaches. *Am. Sci*, 91: 138-149, 2003.
- [11] Marovac J. Investigación y desarrollo de nuevos medicamentos: de la molécula al farmaco. *Revista médica de Chile*, 129 (1): 99-106, 2001.

doi: 10.4067/S0034-98872001000100015

[12] Escalona J, Carrasco R, Padrón J. Introducción al diseño racional de fármacos. Editorial Universitaria, Ciudad de la Habana, Cuba, pp 12-17, 2008. [13] Sainsbury S, Bird L, Rao V, Shepherd S, Stuart D, Hunter W, Ren J. Crystal Structures of Penicillin-Binding Protein-3 from Pseudomonas aeruginosa: Comparison of Native and Antibiotic-Bound Forms. J. Mol. Biol. 405: 173-184, 2011.

doi: 10.1016/j.jmb.2010.10.024

[14] Kong KF, Schneper L, Mathee K. Beta-lactam antibiotics: from antibiosis to resistance and bacteriology. *APMIS* : acta pathologica, microbiologica, et immunologica Scandinavica, 118(1): 1–36, 2010.

doi: 10.1111/j.1600-0463.2009.02563.x

- [15] Protein Data Bank (PDB). Available at: https://www.rcsb.org/structure/4BJP. [Accessed Nobember 6, 2020].
- [16] Sauvage E, Derouaux A, Fraipont C, Joris MHR. et al. Crystal Structure of Penicillin-Binding Protein 3 (PBP3) from Escherichia coli. *PLoS ONE*. 9(5): e98042, 2014.

doi: 10.1371/journal.pone.0098042

[17] Ghuysen JM. Serine beta-lactamases and penicillin-binding proteins. *Annu. Rev. Microbiol.* 45: 37-67, 1991.

doi: 10.1146/annurev.mi.45.100191.000345

[18] Farghaly A, Esmail S, Abdel-Zaher A, Bdel-Hafez A, El-Kashef H. *Bioorg. Med. Chem.* 22: 2166, 2014.

doi: 10.1016/j.bmc.2014.02.019

[19] Salum L, Mascarello A, Canevarolo R, Altei W, Laranjeira A, Neuenfeldt P, Stumpf T. *Eur. J. Med. Chem.* 96: 504, 2015.

doi: 10.1016/j.ejmech.2015.02.041

[20] Hammad SG, El-Gazzar MG, Abutaleb NS, Li D, Ramming I, Shekhar A, Abdel-Halim M, Elrazaz EZ, Seleem MN, Bilitewski U, Abouzid KAM, El-Hossary EM. *Bioorg. Chem.* 95: 103517, 2020.

doi: 10.1016/j.bioorg.2019.103517

- [21] Abo-Ashour MF, Eldehna WM, George RF, Abdel-Aziz MM, Elaasser MM, Gawad NMA, Gupta A, Bhakta S, Abou-Seri SM. *Eur. J. Med. Chem.* 160: 49-60, 2018.
- [22] Rashid M, Husain A, Shaharyar M, Mishra R, Hussain A, Afzal O. Eur. J. Med. Chem. 83: 630, 2014.

doi: 10.1016/j.ejmech.2014.06.033

[23] Kumar D, Singh SP. *Heterocycles*, 63(1): 145, 2004.

doi: 10.3987/REV-03-569

[24] Khunt RC, Khedkar VM, Chawda RS, Chauhan NA, Parikh AR, Coutinho EC. *Bioorg. Med. Chem. Lett.* 22: 666-678, 2012.

doi: 10.1016/j.bmcl.2011.10.059

- [25] Shaquiquzzaman M, Khan S, Amir M, Alam M. Saudi. Pharm. J. 20: 149-154, 2012.
 doi: 10.1016/j.jsps.2011.09.007
- [26] Premnath D, Indiraleka M. Asian. J. Med. Anal. Chem. 1: 27-32, 2014.
- [27] Taj T, Kamble R, Gireesh T, Hunnur R, Margankop S. Eur. J. Med. Chem. 46: 4366-73, 2011.

doi: 10.1016/j.ejmech.2011.07.007

[28] Kumar H, Saini D, Jain S, Jain N. Eur. J. Med. Chem. 70: 248-258, 2013.

doi: 10.1016/j.ejmech.2013.10.004

[29] Spring D. Diversity-oriented synthesis; a challenge for synthetic chemists. *Org. Biomol. Chem.* 1: 3867-3870, 2003.

doi: 10.1039/B310752N

[30] Moreno-Díaz H, Villalobos-Molina R, Ortiz-Andrade R, Díaz-Coutiño D, Medina-Franco JL, Webster SP, et al. *Bioorg. Med. Chem. Lett.* 18: 2871–2877, 2008.

doi: 10.1016/j.bmcl.2008.03.086

[31] Navarrete-Vázquez G, Moreno-Diaz H, Aguirre-Crespo F, León-Rivera I, Villalobos-Molina R, Muñoz-Muñiz O, et al. *Bioorg. Med. Chem. Lett.* 16: 4169-4173, 2006.

doi: 10.1016/j.bmcl.2006.05.082

[32] Cheng F, Li W, Zhou Y, Shen J, Wu Z, Liu G, et al. admetSAR: A Comprehensive Source and Free Tool for Assessment. J. Chem. Inf. Model. 52: 3099-3105, 2012.

doi: 10.1021/ci300367a

- [33] Sashidhara KV, Rosaiah JN, Kumar A. Synth. Commun. 39: 13, 2288-2296, 2009.
 doi: 10.1080/00397910802654724
- [34] Hodge H, Sterner J. Am. Ind. Hyg. Assoc. Q. 10: 94-97, 1949.
- [35] Durst HD, Gokel GW. Química Orgánica Experimental. Reverté S.A., Barcelona, España, pp 325-327, 1985.
- [36] Jawale DV, Pratap UR, Bhosale MR, Mane RA. J. Het. Chem. 53: 1626-1630, 2016.
 doi: 10.1002/jhet.673
- [37] Qiya Z, Hong XH, Suhui W, Shuajiang T, Liangce R. Synth. Commun. 39: 516-522, 2009.
 doi: 10.1080/00397910802399932

Diseño asistido por computador, síntesis y caracterización de híbridos moleculares de dihidropirazoles, aminopirimidinas y tiazolidín-4-onas como inhibidores potenciales de la proteína de unión a penicilina 3 (PBP-3) de *Escherichia coli*.

Resumen: Se utilizó una aproximación asistida por computador para modelar y estudiar andamios heterocíclicos privilegiados que contenían fracciones de dihidropirazol, pirimidina-2-amina y tiazolidín-4-ona (farmacóforos híbridos) para obtener moléculas prototipo antimicrobianas nuevas y promisorias. Se definieron los principales descriptores de biodisponibilidad para determinar la similaridad a fármaco de los compuestos diseñados y para pre-filtrar los once compuestos que exhibían los mejores perfiles, pasando así al estudio de acoplamiento molecular contra una proteína de unión a penicilina tipo 3 clave de *E. coli* enterotoxigénica. Se escogieron siete estructuras por sus energías de afinidad y sus interacciones de acoplamiento con residuos clave en el sitio activo del receptor. Por medio de reacciones de acoplamiento, pertenecientes a las series de chalconas, dihidropirazoles, aminopirimidinas y tiazolidin-4-onas. La elucidación estructural de los productos finales se llevó a cabo por análisis espectral infrarrojo y experimentos de RMN. Estos híbridos moleculares, considerados potenciales hitos en la búsqueda de nuevos compuestos antibacteriales, serán evaluados *in vitro* en estudios posteriores.

Palabras Clave: InhibidoresPBP-3; farmacóforos híbridos; tamizaje *in silico*; dihidropirazoles; pyrimidin-2-aminas; tiazolidín-4-onas.

Desenho assistido por computador, síntese e caracterização de híbridos moleculares de dihidropirazois, aminopirimidinas e tiazolidin-4-onas como potenciais inibidores da proteína de união a penicilina 3 (PBP-3) de *Escherichia coli*.

Resumo: Uma aproximação assistida por computador foi utilizada para modelar e estudar andaimes heterocíclicos privilegiados que continham frações de dihidropirazol, 2-aminopirimidina e tiazolidin-4-ona (farmacóforos híbridos) para obter protótipos de moléculas antimicrobianas novas e promissoras. Definiram-se os principais descritores de biodisponibilidade para determinar a similaridade a fármacos para os compostos desenhados e para selecionar os onze compostos que exibiram os melhores perfis, passando assim ao estudo de acoplamento molecular contra uma proteína chave de união a penicilina tipo 3 de E. coli enterotoxigénica. Foram escolhidas sete estruturas por suas energias de afinidade e suas interações de acoplamento com resíduos chave nos sítios ativos do receptor. Sete compostos com as maiores pontuações de docking foram preparados por meio de reações de condensação ou ciclocondensação, pertencentes às series de chalconas, dihidropirazois, aminopirimidinas e tiazolidin-4-onas. A elucidação estrutural dos produtos finais foi realizada por análise espectral por infravermelho e experimentos de RMN. Tais híbridos moleculares considerados como potenciais acertos na busca por novos compostos antibacterianos serão testados *in vitro* em estudos posteriores.

Palavras-chave: Inibidores PBP-3; farmacóforos híbridos; screening in silico; dihidropirazois; 2-amino-pirimidinas; tiazolidin-4-onas.

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